

# Bacteria Cryopreservation Using a CoolRack® Module

## Introduction

Snap freezing, or flash freezing, is the process by which samples are lowered to temperatures below  $-70^{\circ}\text{C}$  very rapidly using dry ice or liquid nitrogen. Snap freezing achieves the same endpoint as slow rate-controlled freezing, but at a much faster rate ( $-10$  to  $1000^{\circ}\text{C}/\text{minute}$ , as compared to  $-1^{\circ}\text{C}/\text{minute}$ ). Snap freezing with a thermo-conductive CoolRack® tube module will provide sample vessel stability, organized and consistent freezing parameters, rapid hands-free sample processing while avoiding lost or contaminated samples. Snap freezing is performed on a pre-cooled CoolRack module, which ensures fast heat transfer. This method can provide excellent specimen integrity and a wide array of options for analysis, including extraction of proteins, DNA and RNA for use in research and diagnostics.

The following protocol describes a **general procedure** for cryopreserving bacteria for long-term storage. Verify with your laboratory SOP for specific needs for each strain.

## Materials

- Bacterium preparation
- Cryoprotective agent
- CoolBox™ CFT30 ice-free cooling system
- Green CoolBox 30 System freezing cartridge
- CoolRack® CFT30 module
- Cryolabels and/or cryomarkers
- ThermalTray™ LP platform (optional)
- 50 mL reagent reservoirs
- CoolSink® LX55 reagent reservoir holder (optional)
- TruCool® cryogenic vials
- TruCool® hinged cryostorage boxes
- $37^{\circ}\text{C}$  waterbath
- $-80^{\circ}\text{C}$  freezer

## Bacteria Preparation

Follow the laboratory protocol for viral growth and/or purification. Refer to Centers for Disease Control and Prevention (CDC) guidelines for utilization of pathogens in specific Biosafety Level (BSL). *Pathogens are infectious agents and should always be manipulated under a biosafety cabinet with laminar flow.*

## Bacteria Freezing

1. As a general rule, maintain the bacterium preparation at  $4^{\circ}\text{C}$  by placing it in a reagent reservoir and place the reservoir on a thermo-conductive CoolSink LX55 module for uniform and stable cooling. Rest the CoolSink LX55 on ice directly, or on a ThermalTray LP platform for greater stability to minimize ice contact with the reservoir and its contents.
2. Prepare a bacteria glycerol stock by diluting the bacterial preparation with a sterile glycerol solution for a final 15-50% v/v glycerol concentration. Dispense 1 mL of stock (or desired amount) into a pre-labeled TruCool cryogenic vial. To avoid titer reduction maintain the vials at  $4^{\circ}\text{C}$  in a CoolRack CFT30 in the CoolBox. The CoolRack CFT30 module will ensure uniform temperature to all vials and minimizes contamination and spill accidents by allowing one-handed opening/closing of the cryogenic vials.
3. While bacteria samples are kept cold at  $4^{\circ}\text{C}$  in the CoolRack CFT30, equilibrate a second CoolRack CFT30 on dry-ice for 10 minutes. **Note: with this protocol, there is no need to make a dry-ice/ETOH slurry.** Place the vials directly on the pre-equilibrated CoolRack CFT30 module on dry ice and freeze the samples for 3-5 min.
4. Transfer the frozen samples to a TruCool hinged cryostorage box and place it in the  $-80^{\circ}\text{C}$  freezer for long-term storage.

## Bacteria Thawing

1. Place the cryogenic vials from the  $-80^{\circ}\text{C}$  freezer in a CoolBox CFT30 with a pre-frozen green cartridge inside it to maintain the vials at the correct temperature and allow transport of the vials in a safe manner.
2. Place the vials directly in a  $37^{\circ}\text{C}$  water bath, and manually slowly agitate the vials to enable the thawing process. Just before the whole liquid is completely thawed, remove the vial from the  $37^{\circ}\text{C}$  water bath and place it on a CoolRack CFT30 module equilibrated to  $<4^{\circ}\text{C}$  on ice. Samples are ready for experimental procedures or titer assessment.

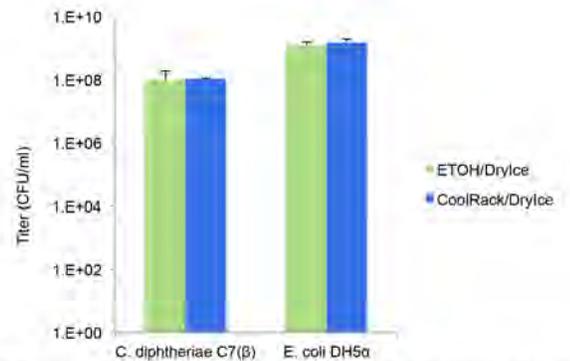


Figure 1: Graph showing the Titer (CFU/ml) of 2 different bacterial strains C.diphtheriae C7 and E. coli DH5 using the 2 freezing methodologies.